


	1990-1991	1991-1992	1992-1993	1993-1994	1994-1995	1995-1996	1996-1997	1997-1998	1998-1999	1999-2000	2000-2001	2001-2002	2002-2003	2003-2004	2004-2005	2005-2006	2006-2007	2007-2008	2008-2009	2009-2010	2010-2011	2011-2012	2012-2013	2013-2014	2014-2015	2015-2016	2016-2017	2017-2018	2018-2019	2019-2020	2020-2021	2021-2022	2022-2023	2023-2024	2024-2025	2025-2026	2026-2027	2027-2028	2028-2029	2029-2030	2030-2031	2031-2032	2032-2033	2033-2034	2034-2035	2035-2036	2036-2037	2037-2038	2038-2039	2039-2040	2040-2041	2041-2042	2042-2043	2043-2044	2044-2045	2045-2046	2046-2047	2047-2048	2048-2049	2049-2050	2050-2051	2051-2052	2052-2053	2053-2054	2054-2055	2055-2056	2056-2057	2057-2058	2058-2059	2059-2060	2060-2061	2061-2062	2062-2063	2063-2064	2064-2065	2065-2066	2066-2067	2067-2068	2068-2069	2069-2070	2070-2071	2071-2072	2072-2073	2073-2074	2074-2075	2075-2076	2076-2077	2077-2078	2078-2079	2079-2080	2080-2081	2081-2082	2082-2083	2083-2084	2084-2085	2085-2086	2086-2087	2087-2088	2088-2089	2089-2090	2090-2091	2091-2092	2092-2093	2093-2094	2094-2095	2095-2096	2096-2097	2097-2098	2098-2099	2099-2100	2100-2101	2101-2102	2102-2103	2103-2104	2104-2105	2105-2106	2106-2107	2107-2108	2108-2109	2109-2110	2110-2111	2111-2112	2112-2113	2113-2114	2114-2115	2115-2116	2116-2117	2117-2118	2118-2119	2119-2120	2120-2121	2121-2122	2122-2123	2123-2124	2124-2125	2125-2126	2126-2127	2127-2128	2128-2129	2129-2130	2130-2131	2131-2132	2132-2133	2133-2134	2134-2135	2135-2136	2136-2137	2137-2138	2138-2139	2139-2140	2140-2141	2141-2142	2142-2143	2143-2144	2144-2145	2145-2146	2146-2147	2147-2148	2148-2149	2149-2150	2150-2151	2151-2152	2152-2153	2153-2154	2154-2155	2155-2156	2156-2157	2157-2158	2158-2159	2159-2160	2160-2161	2161-2162	2162-2163	2163-2164	2164-2165	2165-2166	2166-2167	2167-2168	2168-2169	2169-2170	2170-2171	2171-2172	2172-2173	2173-2174	2174-2175	2175-2176	2176-2177	2177-2178	2178-2179	2179-2180	2180-2181	2181-2182	2182-2183	2183-2184	2184-2185	2185-2186	2186-2187	2187-2188	2188-2189	2189-2190	2190-2191	2191-2192	2192-2193	2193-2194	2194-2195	2195-2196	2196-2197	2197-2198	2198-2199	2199-2200	2200-2201	2201-2202	2202-2203	2203-2204	2204-2205	2205-2206	2206-2207	2207-2208	2208-2209	2209-2210	2210-2211	2211-2212	2212-2213	2213-2214	2214-2215	2215-2216	2216-2217	2217-2218	2218-2219	2219-2220	2220-2221	2221-2222	2222-2223	2223-2224	2224-2225	2225-2226	2226-2227	2227-2228	2228-2229	2229-2230	2230-2231	2231-2232	2232-2233	2233-2234	2234-2235	2235-2236	2236-2237	2237-2238	2238-2239	2239-2240	2240-2241	2241-2242	2242-2243	2243-2244	2244-2245	2245-2246	2246-2247	2247-2248	2248-2249	2249-2250	2250-2251	2251-2252	2252-2253	2253-2254	2254-2255	2255-2256	2256-2257	2257-2258	2258-2259	2259-2260	2260-2261	2261-2262	2262-2263	2263-2264	2264-2265	2265-2266	2266-2267	2267-2268	2268-2269	2269-2270	2270-2271	2271-2272	2272-2273	2273-2274	2274-2275	2275-2276	2276-2277	2277-2278	2278-2279	2279-2280	2280-2281	2281
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Jonathan L. Klein Reg. No. 41,119
Attorney for Applicants

JLK/SA/ba

[Figure 5 is] Figs. 5A and 5B, collectively and consecutively, show a partial cDNA sequence SEQ ID NO:8 and the corresponding deduced amino acid sequence SEQ ID NO:9 of a colon specific gene of the present invention.

Page 6, third full paragraph, which begins "Figure 6 is...":

Figure 6 is a partial cDNA sequence SEQ ID NO:10 and the corresponding deduced amino acid sequence SEQ ID NO:11 of a colon specific gene of the present invention.

Page 6, fourth full paragraph, which begins "Figure 7 is...":

Figure 7 is a partial cDNA sequence SEQ ID NO:12 of a colon specific gene of the present invention.

Page 6, fifth full paragraph, which begins "Figure 8 is...":

Figure 8 is a full length cDNA sequence SEQ ID NO:13 and the corresponding deduced amino acid sequence SEQ ID NO:14 of a colon specific gene of the present invention.

Page 6, sixth full paragraph, which begins "Figure 9 is...":

[Figure 9 is] Figs. 9A and 9B, collectively and consecutively, show a full length cDNA sequence SEQ ID NO:15 and corresponding deduced amino acid sequence SEQ ID NO:16 of [a] the CSG10 colon specific gene of the present invention.

Page 6, seventh full paragraph, which begins "Figure 10 is...":

Figure 10 is a partial cDNA sequence SEQ ID NO:17 and corresponding deduced amino acid sequence SEQ ID NO:18 of a colon specific gene of the present invention.

Page 6, eighth full paragraph, which begins "Figure 11 is...":

Figure 11 is a partial cDNA sequence SEQ ID NO:19 and the corresponding deduced amino acid sequence SEQ ID NO:20 of a colon specific gene of the present invention.

Page 6, ninth full paragraph, which begins "Figure 12 is...":

Figure 12 is a partial cDNA sequence SEQ ID NO:21 of a colon specific gene of the present invention.

Page 6, tenth full paragraph, which begins "Figure 13 is...":

Figure 13 is a partial cDNA sequence SEQ ID NO:22 of a colon specific gene of the present invention.

Page 6, twelfth full paragraph extending onto page 7, which begins "In accordance with one aspect...":

In accordance with one aspect of the present invention there is provided a polynucleotide which encodes one of the mature polypeptides having the deduced amino acid sequence of [Figure 8 or 9] Fig. 8 or of Figs. 9A and 9B collectively, and fragments, analogues and derivatives thereof.

At page 7, first full paragraph, which begins "In accordance with a further aspect...":

In accordance with a further aspect of the present invention there is provided a polynucleotide which encodes the same mature polypeptide as a human gene having a coding portion which contains a polynucleotide which is at least 90% identical (preferably at least 95% identical and most preferably at least 97% or 100% identical) to one of the polynucleotides of [Figures 1-7 or 9-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-7, 9A-9B and 10-13, as well as fragments thereof.

Page 7, third full paragraph, which begins "In accordance with yet another aspect...", and replace therewith the following paragraph:

In accordance with yet another aspect of the present invention, there is provided a polynucleotide probe which hybridizes to mRNA (or the corresponding cDNA) which is transcribed from the coding portion of a human gene which coding portion includes a DNA sequence which is at least 90% identical to (preferably at least 95% identical to) and most preferably at least 97% or 100% identical) to one of the polynucleotide sequences of [Figures 1-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-8, 9A-9B and 10-13.

which coding sequence includes the DNA of [Figures 1-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-8, 9A-9B and 10-13 or the deposited cDNA.

Page 9, fourth full paragraph, which begins "The polynucleotides of the invention...":

The polynucleotides of the invention may have a coding sequence which is a naturally occurring allelic variant of the human gene whose coding sequence includes DNA as shown in [Figures 1-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-8, 9A-9B and 10-13 or of the coding sequence of the DNA in the deposited clone. As known in the art, an allelic variant is an alternate form of a polynucleotide sequence which may have a substitution, deletion or addition of one or more nucleotides, which does not substantially alter the function of the encoded polypeptide.

Page 10, third full paragraph extending onto page 10, which begins "The present invention further relates...":

The present invention further relates to polynucleotides which hybridize to the hereinabove-described polynucleotides if there is at least 70%, preferably at least 90%, and more preferably at least 95% identity between the sequences. The present invention particularly relates to polynucleotides which hybridize under stringent conditions to the hereinabove-described polynucleotides. As herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences. The polynucleotides which hybridize to the hereinabove described polynucleotides in a preferred embodiment encode polypeptides which either retain substantially the same biological function or activity as the mature polypeptide of the present invention encoded by a coding sequence which includes the DNA of [Figures 1-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-8, 9A-9B and 10-13 or the deposited cDNA(s).

Page 11, second full paragraph, which begins "Thus, the present invention is...":

Thus, the present invention is directed to polynucleotides having at least a 70% identity, preferably at least 90% and more preferably at least 95% identity to a polynucleotide which encodes the mature polypeptide encoded by a human gene which

includes the DNA of one of [Figures 1-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-8, 9A-9B and 10-13 as well as fragments thereof, which fragments have at least 30 bases and preferably at least 50 bases and to polypeptides encoded by such polynucleotides.

At page 11, please delete the fourth full paragraph extending onto page 12, which begins "The partial sequences of Figures 1-7...", and replace therewith the following paragraph:

The partial sequences of [Figures 1-7 and 10-13] Figs. 1, 2A-2B, 3A-3B, 4, 5A-5B, 6-7 and 10-13 may be used to identify the corresponding full length gene from which they were derived. The partial sequences can be nick-translated or end-labelled with ^{32}P using polynucleotide kinase using labelling methods known to those with skill in the art (Basic Methods in Molecular Biology, L.G. Davis, M.D. Dibner, and J.F. Battey, ed., Elsevier Press, NY, 1986). A lambda library prepared from human colon tissue can be directly screened with the labelled sequences of interest or the library can be converted en masse to pBluescript (Stratagene Cloning Systems, La Jolla, CA 92037) to facilitate bacterial colony screening. Regarding pBluescript, see Sambrook et al., Molecular Cloning-A Laboratory Manual, Cold Spring Harbor Laboratory Press (1989), pg. 1.20. Both methods are well known in the art. Briefly, filters with bacterial colonies containing the library in pBluescript or bacterial lawns containing lambda plaques are denatured and the DNA is fixed to the filters. The filters are hybridized with the labelled probe using hybridization conditions described by Davis et al., supra. The partial sequences, cloned into lambda or pBluescript, can be used as positive controls to assess background binding and to adjust the hybridization and washing stringencies necessary for accurate clone identification. The resulting autoradiograms are compared to duplicate plates of colonies or plaques; each exposed spot corresponds to a positive colony or plaque. The colonies or plaques are selected, expanded and the DNA is isolated from the colonies for further analysis and sequencing.

At page 21, third full paragraph, which begins "The polypeptides of the present invention...", and replace therewith the following paragraph:

The polypeptides of the present invention include the polypeptides of [Figures 8 and 9] Fig. 8 and of Figs. 9A and 9B, collectively, (in particular the mature polypeptides)

as well as polypeptides which have at least 70% similarity (preferably at least a 70% identity) to the polypeptides of [Figures 8 and 9] Fig. 8 and of Figs. 9A and 9B, collectively, and more preferably at least a 90% similarity (more preferably at least a 90% identity) to the polypeptides of [Figures 8 and 9] Fig. 8 and of Figs. 9A and 9B, collectively, and still more preferably at least a 95% similarity (still more preferably at least 95% identity) to the polypeptides of [Figures 8 and 9] Fig. 8 and of Figs. 9A and 9B, collectively, and also include portions of such polypeptides with such portion of the polypeptide generally containing at least 30 amino acids and more preferably at least 50 amino acids.

Page 43, second full paragraph extending onto pages 44 and 45, which begins "The DNA sequence encoding a polypeptide...":

The DNA sequence encoding a polypeptide of the present invention, ATCC # [97201] 97102, which one is initially amplified using PCR oligonucleotide primers corresponding to the 5' sequences of the processed protein (minus the signal peptide sequence) and the vector sequences 3' to the gene. Additional nucleotides corresponding to the DNA sequence are added to the 5' and 3' sequences respectively. The 5' oligonucleotide primer may contain, for example, a restriction enzyme site followed by nucleotides of coding sequence starting from the presumed terminal amino acid of the processed protein. The 3' sequence may, for example, contain complementary sequences to a restriction enzyme site and also be followed by nucleotides of the nucleic acid sequence encoding the protein of interest. The restriction enzyme sites correspond to the restriction enzyme sites on a bacterial expression vector, for example, pQE-9 (Qiagen, Inc. Chatsworth, CA). pQE-9 encodes antibiotic resistance (Amp^r), a bacterial origin of replication (ori), an IPTG-regulatable promoter operator (P/O), a ribosome binding site (RBS), a 6-His tag and restriction enzyme sites. pQE-9 is then digested with the restriction enzymes corresponding to restriction enzyme sites contained in the primer sequences. The amplified sequences are ligated into pQE-9 and inserted in frame with the sequence encoding for the histidine tag and the RBS. The ligation mixture is then used to transform an E. coli strain, for example, M15/rep 4 (Qiagen) by the procedure described in Sambrook, J. et al., Molecular Cloning: A Laboratory Manual, Cold Spring Laboratory Press, (1989). M15/rep4 contains multiple copies of the plasmid pREP4, which expresses the lacI repressor and also confers kanamycin resistance (Kan^r). Transformants are

identified by their ability to grow on LB plates and ampicillin/kanamycin resistant colonies are selected. Plasmid DNA is isolated and confirmed by restriction analysis. Clones containing the desired constructs are grown overnight (O/N) in liquid culture in LB media supplemented with both Amp (100 ug/ml) and Kan (25 ug/ml). The O/N culture is used to inoculate a large culture at a ratio of 1:100 to 1:250. The cells are grown to an optical density 600 (O.D.⁶⁰⁰) of between 0.4 and 0.6. IPTG ("Isopropyl-B-D-thiogalactopyranoside") is then added to a final concentration of 1 mM. IPTG induces by inactivating the lacI repressor, clearing the P/O leading to increased gene expression. Cells are grown an extra 3 to 4 hours. Cells are then harvested by centrifugation. The cell pellet is solubilized in the chaotropic agent 6 Molar Guanidine HCl. After clarification, solubilized protein is purified from this solution by chromatography on a Nickel-Chelate column under conditions that allow for tight binding by proteins containing the 6-His tag (Hochuli, E. et al., J. Chromatography 411:177-184 (1984)). The protein is eluted from the column in 6 molar guanidine HCl pH 5.0 and for the purpose of renaturation adjusted to 3 molar guanidine HCl, 100mM sodium phosphate, 10 mmolar glutathione (reduced) and 2 mmolar glutathione (oxidized). After incubation in this solution for 12 hours the protein is dialyzed to 10 mmolar sodium phosphate.

Page 47, first full paragraph, which begins "Preparation of templates by PCR...":

Preparation of templates by PCR is a modification of the method of Rosenthal et al. (Rosenthal, et al., Nucleic Acids Res., 1993, 21:173-174). Colonies containing cDNA cloned into pBluescript II or rescued pBluescript phagemid are grown overnight in LB containing ampicillin in a 96 well tissue culture plate. Two µl of the cultures are used as template in a PCR reaction (Saiki, RK, et al., Science, 239:487-493, 1988; and Saiki, RK, et al., Science, 230:1350-1354, 1985) using a tricine buffer system (Ponce and Micol., Nucleic Acids Res., 1992, 20:1992.) and 200 µM dNTPs. The primer set chosen for amplification of the templates is outside of primer sites chosen for sequencing of the templates. The primers used are 5'-ATGCTTCCGGCTCGTATG-3' (SEQ ID NO:23) which is 5' of the M13 reverse sequence in pBluescript and 5'-GGGTTTTCACGTCACGAC-3' (SEQ ID NO:24), which is 3' of the M13 forward primer in pBluescript. Any primers which correspond to the sequence flanking the M13

[illegible]